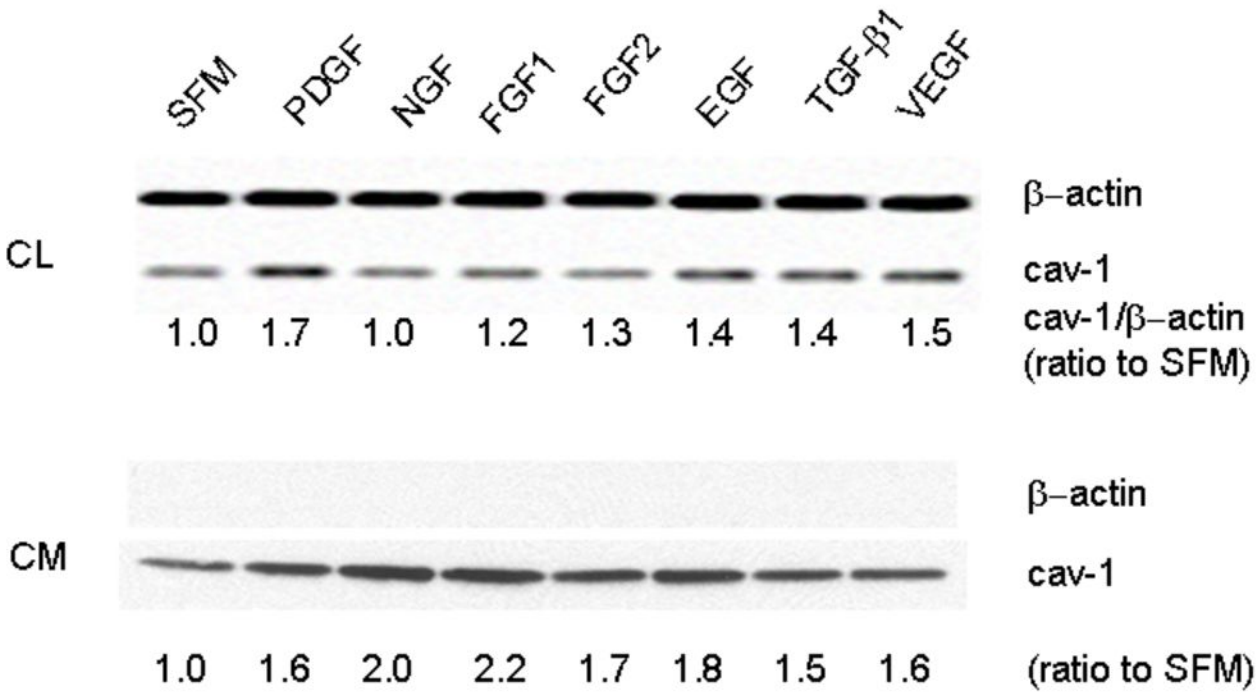
Supplemental Fig. 1



**Supplemental Materials and Methods**

*Induction of Cav-1 Expression and Secretion by GFs*

ABAC3 cells were seeded in complete culture medium and grown overnight. After being rinsed once with serum-free medium (SFM), cells were incubated in SFM with or without a specific GF at the following concentrations: human GFs: platelet-derived GF (PDGF)-AB, 10 ng/mL (Invitrogen); never GF (NGF), 10 ng/mL (Prospec); FGF1, 5 ng/mL (Invitrogen); FGF2, 2 ng/mL (Invitrogen); epidermal GF (EGF), 5 ng/mL (Invitrogen); TGF-β1, 1 ng/mL (Invitrogen), and VEGF, 10 ng/mL (Upstate). Mouse GFs: PDGF-BB, 10 ng/mL (Invitrogen); NGF, 10 ng/mL (Invitrogen); FGF1, 5 ng/mL (R & D Systems); FGF2, 2 ng/mL (Invitrogen); EGF, 5 ng/mL (Invitrogen); and VEGF, 10 ng/mL (Invitrogen).

For determining cav-1 expression, cell lysates were prepared 48 h after treatment with the GFs. For determining secreted cav-1 levels, conditioned media were collected 24 h after treatment with the GFs. The conditioned media were then centrifuged once at 130 $\times$ *g* for 5 min to remove floating cells and once at 10,000 $\times$ *g* for 20 min to removed remaining insoluble materials. The resulting supernatants were concentrated 50 to 100 fold using Amicon Ultra-4 with 10-kDa cutoff. The equivalents of conditioned media produced from $5.0 \times 10^5$ cells were loaded on gel for the determination of secreted cav-1 levels.

Quantitative analysis were performed by measuring the density of each protein bands using a computer-assisted software (Nikon, NIS-Elements AR3.0) and by making calculations according to the following methods: for cell lysates, the protein bands of interest were first normalized by corresponding β–actin and each nomorlized value was then compared to control (control as 1); for conditioned medium, the protein bands of interest were compared to control (SFM).