Interleukin-6 and Oncostatin-M Synergize with the PI3K/AKT Pathway to Promote Aggressive Prostate Malignancy in Mouse and Human Tissues

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Abstract

Chronic inflammation has been proposed as an etiological and progression factor in prostate cancer. In this study, we used a dissociated prostate tissue recombination system to interrogate the role of interleukin 6 (IL6) and the related cytokine oncostatin-M (OSM) in the initiation and progression of prostate cancer. We identified that prostatic intraepithelial neoplasia (PIN) lesions induced by PTEN loss of function (PTENLOF) progress to invasive adenocarcinoma following paracrine expression of either cytokine. Increased expression of OSM was also able to drive progression of benign human epithelium when combined with constitutively activated AKT. Malignant progression in the mouse was associated with invasion into the surrounding mesenchyme and increased activation of STAT3 in PTENLOF grafts expressing IL6 or OSM. Collectively, our work indicates that pro-inflammatory cytokines such as IL6 or OSM could activate pathways associated with prostate cancer progression and synergize with cell-autonomous oncogenic events to promote aggressive malignancy.

Implications: Increased expression of IL6 or OSM synergizes with loss of PTEN to promote invasive prostate cancer.

Visual Overview: http://mcr.aacrjournals.org/content/early/2013/09/02/1541-7786.MCR-13-0238/F1.large.jpg.

Introduction

Chronic inflammation is observed in nearly all forms of cancer and may represent a contributing factor at initiation, progression, and metastasis (1). Numerous factors have been proposed to contribute to prostatic inflammation and it has also been proposed to play a role driving in the multifocal nature of prostate cancer (2). Studies using bacterial colonization of the prostate to elicit chronic inflammatory conditions have observed epithelial reactive hyperplasia and prostatic intraepithelial neoplasia (PIN) lesions (3). Increased expression of inflammatory cytokines, including members of the interleukin 6 (IL6) family, has been observed in these models and could represent a functional mechanism (4). Immunohistochemical analysis of human prostate biopsies have shown increased expression of IL6 and the related OSM ligands and their cognate receptors with prostate cancer progression (5). Therefore, we sought to interrogate the role of inflammation by monitoring transformation of prostate epithelial cells in a pro-inflammatory environment induced by ectopic expression of IL6 or the related OSM in the stroma of tissue recombination experiments.

The IL6 family of cytokines consists of several related ligands that share common protein structure and use of GP130 signal transducer with receptor adaptors modulating ligand specificity (6). Studies have identified increased serum levels of IL6 in patients with prostate cancer correlating to increased clinical stage, hormone-refractory disease, and metastasis (7, 8). Several prostate cancer cell lines have been shown to express IL6 whereas treatment with either cytokine generally confers a growth advantage (9). Antibodies targeting IL6 inhibit xenograft growth and castration resistance, although they have shown poor clinical efficacy to date, indicating the need to more fully understand how these molecules function in the tumor microenvironment (11, 12).
We sought to interrogate the functional roles of IL6 and OSM in prostate cancer in vivo using a dissociated tissue recombination system developed in our laboratory (13, 14). We identified that paracrine expression of either IL6 or OSM is sufficient to promote invasive progression from intermediate PIN lesions induced by PTENΔLOF in mouse tissues or activation of AKT in human tissues. Increased expression of either IL6 or OSM in the context of PTEN loss resulted in increased activation of STAT3 over PTEN loss alone, indicating a potential molecular mechanism. That cytokines related to IL6 exhibit similar cancer progression behavior indicates that highly targeted therapies aimed solely at disrupting IL6 signaling could be less effective than those targeting signaling nodes common to the IL6 family.

Materials and Methods

Mouse strains and regeneration assay

Housing, maintenance, surgical, and experimental procedures were undertaken in compliance with the regulations of the Division of Laboratory Animal Medicine of the University of California, Los Angeles (Los Angeles, CA). Homozygous Pten Conditional Knockout (PtenΔLOF), strain B6.129S4-Pten<sup>fl/fl</sup>/Pten<sup>tm1Hwu/J</sup>, were purchased from Jackson Laboratory and maintained in our facility. Lentivirus preparation, titration, and infection, and regeneration of dissociated cells were conducted as previously described under safety regulations for lentivirus use set by Environmental Health and Safety (EHE&S) at University of California, Los Angeles. Briefly, prostate tissue from 6- to 12-week-old male mice was minced, digested, and dissociated according to published protocols (13). Dissociated cells (1 × 10<sup>5</sup> to 2 × 10<sup>5</sup>) were infected with lentivirus at a multiplicity of infection (MOI) of 50, recombined with UGSM (1 × 10<sup>5</sup> to 2 × 10<sup>5</sup>), suspended in a collagen plug, and engrafted under the kidney capsule for 6 to 10 weeks. Grafts were recovered via surgical resection of the kidney and fixed in 10% buffered formalin overnight or flash frozen. UGSM cells were prepared from pregnant C57BL/6 females on embryonic day 14 and cultured in UGSM media [Dulbecco's Modified Eagles Media (DMEM), 5% FBS, 5% NuSerum (BD #355504), 1 × selenium-transferrin-insulin (Gibco #51500-056), 2 mmol/L l-glutamine].

Human prostate regenerations

For primary human cells, we used a protocol approved through the UCLA Office for the Protection of Research Subjects, and all human tissue samples were de-identified to protect patient confidentiality. A total of 3 patient samples were used for this study and all specimens were processed as described previously (14). Briefly, surgical prostate specimens were removed and cancerous areas were separated from the benign tissue. Basal epithelial cells were isolated by fluorescence-activated cell sorting (FACS) from dissociated benign tissue using phycoerythrin (PE)-conjugated CD49f (eBiosciences #12-0495-83) and allophycocyanin (APC)-conjugated Trop2 (FAB650A, R&D Systems) antibodies. Cells were stained in PrEGM supplemented with 2.5 μg/mL Fungizone (Gibco) and 10 μmol/L of the p160 ROCK inhibitor Y-27632 dihydrochloride (Tocris Bioscience #1254). Sorting was conducted on a BD FACS Aria II (BD Biosciences).

Cell lines and cytokine treatment

PEB cells were a kind gift from Dr. Lynnette Wilson (NYU) maintained in PrEGM (Lonza, # CC-4177 and CC-3165) supplemented with 10% FBS. CaP8 cells were a kind gift from Dr. Hong Wu (UCLA) and maintained in UGSM media. Cell lines were not authenticated. Human recombinant IL6 and OSM were purchased from Cell Signaling (#8904SF and #5367SF, respectively) and were reconstituted in 1× PBS supplemented with 1% bovine serum albumin (BSA). For cytokine treatment assays, cells were serum starved in DMEM supplemented with 0.2% FBS and 2 mmol/L l-glutamine for 6 to 18 hours and treated with cytokines for indicated times. Results presented are representative of 2 replicates.

Results

IL6 and oncostatin-M expression promote progression to invasive prostate cancer in mouse and human tissues

To assess the potential role of IL6 and oncostatin-M (OSM) transformation of both mouse and human prostate epithelium, we used a dissociated prostate regeneration system developed in our laboratory (13, 14). For mouse tissues, we used dissociated prostate cells from adult male homozygous Pten conditional knockout mice transduced with either GFP control or Cre-GFP lentivirus that were combined with urogenital sinus mesenchyme (UGSM) transduced with RFP control, IL6-RFP, or OSM-RFP lentivirus (Fig. 1A). Western blot analysis was used to confirm increased expression and activity of secreted IL6 and OSM in lysates and conditioned media of transduced UGSM (Fig. 1B).

Control grafts exhibited regeneration of normal prostate cellular architecture, whereas PTENΔLOF combined with control UGSM resulted in PIN lesion formation (Fig. 1C). PTENΔLOF combined with UGSM expressing either IL6 or OSM exhibited a heterogeneous range of transformation states, although graft wet weights were not significantly different (Supplementary Fig. S1A–S1C and Supplementary Table S1). IL6-expressing grafts largely exhibited high-grade PIN lesions and locally invasive growth, with a few poorly differentiated lesions present. OSM-expressing grafts often presented as locally invasive lesions with several regions of poorly differentiated adenocarcinoma with anaplastic features, indicating that OSM could promote a more aggressive disease (Supplementary Fig. S1C). Grafts displayed immunohistochemical features of human prostate cancer including nuclear AR expression and loss of P63-expressing basal cells in high-grade lesions (Fig. 1D and Supplementary Fig. S1D). Phenotypically similar results were achieved using autocrine expression of OSM in donor prostate tissue with activated AKT, although grafts with IL6 were significantly inhibited and transformation was not observed (Supplementary Fig. S2). We used immunofluorescent confocal microscopy to qualitatively assess epithelial invasion, using E-
cadherin to identify epithelial cells with respect to the basement membrane component collagen IV. Control and PTEN\textsuperscript{LOF} grafts exhibited normal E-cadherin localization to the basement membrane, whereas IL6 and OSM grafts exhibited disorganized E-cadherin expression with epithelial cells invading into the surrounding mesenchyme (Supplementary Fig. S3).

For human experiments, expression of constitutively active AKT was used as a surrogate for PTEN\textsuperscript{LOF}. Benign human epithelium was isolated from radical prostatectomy samples and transduced with either RFP or AKT-RFP lentivirus. Transduced cells were then combined with UGSM transduced with GFP, IL6-, or OSM-GFP lentivirus, suspended in Matrigel, and injected subcutaneously into immunodeficient hosts (Fig. 2A). Control grafts exhibited normal epithelial regeneration with nuclear AR expression in luminal epithelium and P63 expression in basal cells (Fig. 2B and C and Supplementary Fig. S4A). Expression of activated AKT resulted in PIN lesions similar to those observed in mouse PTEN-knockout regenerations (Fig. 2B). Paracrine expression of either IL6 or OSM alone dramatically inhibited epithelial regeneration with only a few small nests of epithelial cells primarily composed of P63-expressing basal cells (Supplementary Fig. S4B). Similarly, paracrine expression of IL6 in grafts with AKT-infected epithelium also exhibited significant inhibition and no epithelial regeneration or transformation was observed (Fig. 2B and C). Grafts expressing OSM with AKT exhibited small nests of dysplastic epithelial cells that express AR, although they retained P63-expressing cells, indicating the lesions had not progressed to clinical adenocarcinoma (Fig. 2B and C). Activation and overexpression of AKT was confirmed by immunohistochemical analysis for phospho-serine 473 AKT levels and total protein (Fig. 2C and Supplementary Fig. S4C). The quantification of observed phenotypes is reported in Supplementary Table S2.

**Invasive tumor foci identified in grafts expressing IL6 or OSM with loss of PTEN exhibited increased activation of JAK/STAT pathway**

Ligand engagement of IL6 family members activates constitutively Janus-activated kinases (JAK), resulting in activation of the signal transduction and transcription factor (STAT), mitogen-activated protein kinases (MAPK), and AKT pathways (6). To interrogate whether IL6 and OSM exhibit differential activation of downstream pathways, we...
treated benign PEB and tumorigenic CaP8 cell lines with human recombinant IL6 and OSM and assessed activation of downstream pathways by western blot analysis (Fig. 3A). Treatment with IL6 or OSM exhibited increased STAT3 and AKT activation in both PEB and CaP8 cell lines compared with controls, with OSM exhibiting a greater increase over IL6 for both signaling pathways. Neither IL6 nor OSM exhibited a dramatic increase in extracellular signal-regulated kinase 1/2 (ERK1/2) activation over carrier-treated cells.

We used immunohistochemical analysis to interrogate activation of pathways downstream of IL6 and OSM from in vivo transformations using mouse tissues (Fig. 3B). Control regenerations exhibited little-to-no activation of AKT, extracellular signal–regulated kinase ERK1/2, or STAT3. Increased AKT activation was observed in all PTENLOF lesions and were similar across PTENLOF alone and with IL6 or OSM. PTENLOF resulted in increased ERK1/2 activation over control grafts with similar levels observed in grafts from PTENLOF alone with IL6, whereas PTENLOF grafts with OSM exhibited mild, although consistently increased ERK1/2 activation. PTENLOF resulted in increased STAT3 activation compared with control grafts with further increased levels observed in grafts from both PTENLOF with IL6 or OSM. Immunohistochemical analysis for total proteins confirmed basal expression levels in control grafts with all PTENLOF grafts exhibiting increased expression of total AKT, ERK1/2, and STAT3 protein (Supplementary Fig. S5). These results indicate that tumor progression via IL6 and OSM is associated with increased activation of the STAT pathway, although OSM synergy could be also be mediated, in part, by MAPK.

Discussion

Correlative and in vitro data strongly indicate that chronic inflammation could act as an etiologic and progression factor, highlighting the need for further validation using in vivo models. Our study identified that both IL6 and OSM exhibited synergy with PTENLOF and display heterogeneous transformation phenotypes ranging from high-grade PIN and locally invasive lesions to poorly differentiated adenocarcinoma with anaplastic features. We speculate that this complexity is a result of variations in the local concentration...
of IL6 or OSM and could serve as a model for the heterogeneity commonly observed in human prostate cancer. While our work focused on the interaction of IL6 and OSM in the context of PTENLOF, other studies indicate that this synergy could extend to other oncogenic insults. Exogenous expression of IL6 has been shown to transform a nontumorigenic prostate epithelial cell line immortalized with SV40 large T-antigen, indicating potential synergy with

Figure 3. Increased expression of OSM in PTENLOF grafts results in increased phosphorylation of STAT3 and ERK1/2 downstream of IL6 and OSM. A, Western blot analysis of PEB and CaP8 cells treated with carrier or 10 ng/mL of IL6 or OSM in DMEM with 0.2% FBS for indicated times. Treatment with carrier resulted in transient activation of ERK1/2 in both cell lines with a slight increase in AKT and STAT3 activation. Treatment with either IL6 or OSM exhibited activation of both AKT and STAT3 pathways with OSM consistently exhibiting increased activation over IL6. Activation of ERK1/2 above background was not consistently observed. B, immunohistochemical (IHC) analysis of AKT, ERK1/2, and STAT3 activation from normal, PTENLOF, and PTENLOF with IL6 or OSM. All grafts with PTENLOF exhibited increased activation of AKT, ERK1/2, and STAT3 above basal levels in normal regenerations. PTENLOF grafts with IL6 exhibited increased levels of STAT3 phosphorylation over PTENLOF alone with no discernible increase in ERK1/2 activation. PTENLOF with OSM exhibited consistently higher levels of STAT3 phosphorylation with a mild increase in ERK1/2 phosphorylation. Scale bars: 20×, 100 μm; 63×, 50 μm.
inhibition of the TP53 and RB pathways (15). Work by Kan and colleagues identified that while OSM suppressed the growth of normal human breast epithelium, co-expression of cMYC abrogated growth arrest and promoted transformation (16). Our results support this finding as neither cytokine was sufficient to transform mouse or human prostate epithelium on its own in our system. Oncogenic response to IL6 and related cytokines therefore seems to be dependent on the presence coincident mutations within the prostate.

Expression of either IL6 or OSM alone strongly inhibited epithelial regeneration in both mouse and human tissues, whereas studies using prostate cancer cell lines have shown paradoxical effects of either growth promotion or inhibition depending on the cell line. Growth inhibition by IL6-type cytokines in tumor cell lines occurs through a variety of mechanisms including activation of the P27/KIP1 checkpoint (17). A study from Duijn and Trapman indicated that the PI3K/akt pathway modulated expression of SKP2, which mediated the downregulation of P27/KIP1 in DU145 and PC3 but not LNCaP cells (18). Intriguingly, this corresponds with published data concerning prostate cancer cell line response to IL6 (9). Activation of the PI3K/akt pathway by IL6 or OSM in DU145 and PC3 could inhibit the P27/KIP1 checkpoint and promote cell growth, whereas LNCaP cells do not exhibit this same behavior and thus respond by growth inhibition and differentiation. PTENLOF has been associated with increased levels of SKP2 and decreased levels of P27/KIP1 in human prostate cancer biopsy specimens, indicating that this checkpoint might be bypassed in the context of chronic inflammation with PTENLOF, allowing for increased tumor growth (19).

Sequencing studies indicate that prostate cancer does not exhibit dramatic mutation rates compared to many other cancers (20). Studies from our laboratory and others have shown that most single oncogenes are not sufficient to induce invasive prostate cancer, indicating that it is necessary to activate multiple signaling pathways (14). Both IL6 and OSM have been shown to activate STAT3, AKT, and MAPK in several cell types, whereas our in vivo studies suggest strong activation of STAT3 with a mild increase in ERK1/2 (6). These pathways have been implicated in nearly all cancers to some degree and several exhibit functional synergy when co-activated. We propose that chronic inflammation could expose tumor cells to a diverse array of cytokines that could activate multiple oncogenic pathways and serve as a surrogate for direct mutation.

Current therapeutic research strategies are increasingly using highly targeted drugs such as antibody-based therapies designed to inhibit the IL6 ligand or the IL6Ra subunit. The humanized antibody CNOT328 inhibits the conversion to androgen-independent disease and modulates activation of downstream pathways in prostate cancer xenografts, although it has shown poor performance in clinical trials (11, 12). Our study indicates that other members of the IL6 family, and likely other cytokines, can also exhibit pro-tumorigenic functions that can even exceed those of IL6. Highly targeted therapies directed at single ligands such as IL6 could exhibit reduced efficacy due to coexpression of family members with redundant activity. Therapies targeting signaling nodes such as the JAK family could therefore exhibit greater efficacy through inhibition of both related family members and shared downstream pathways such as STAT3 and ERK1/2. Therapeutic strategies such as this would benefit greatly from an increased understanding of how cytokine expression profiles can be used as diagnostic biomarkers.

Disclosure of Potential Conflicts of Interest
No potential conflicts of interest were disclosed.

Authors’ Contributions
Conception and design: D.A. Smith, A. Kiba, Y. Zong, O.N. Witte
Development of methodology: D.A. Smith, A. Kiba, Y. Zong, O.N. Witte
Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): D.A. Smith, A. Kiba, Y. Zong
Analysis and interpretation of data (e.g., statistical analysis, bioinformatics, computational analysis): D.A. Smith, A. Kiba, Y. Zong, O.N. Witte
Writing, review and/or revision of the manuscript: D.A. Smith, A. Kiba, Y. Zong, O.N. Witte
Study supervision: O.N. Witte

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References


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