Fhit Regulates EMT Targets through an EGFR/Src/ERK/Slug Signaling Axis in Human Bronchial Cells

Audrey Joannes,1* Simon Grelet,1* Laurent Duca,2 Christine Gilles,3 Claire Kileztky,1 Véronique Dalstein,1,4 Philippe Birembaut,1,4 Myriam Polette,1,4 Béatrice Nawrocki-Raby1

Authors’ affiliations:

1INSERM UMR-S 903, SFR CAP-Santé (FED 4231), University of Reims Champagne-Ardenne, Reims, F-51100 France
2CNRS FRE 3481, SFR CAP-Santé (FED 4231), University of Reims Champagne-Ardenne, Reims, F-51100 France
3Laboratory of Developmental and Tumor Biology, GIGA-Cancer, University of Liège, CHU of Sart-Tilman, Liège, B-4000 Belgium
4Laboratory of Histology, CHU of Reims, Reims, F-51100 France

*These authors contributed equally to this work

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**Corresponding author:** Béatrice Nawrocki-Raby, INSERM UMR-S 903, CHU Maison Blanche, 45 rue Cognacq-Jay, F-51100 REIMS, France. Phone: 33-3-26-78-77-70; Fax: 33-3-26-06-58-61; E-mail: beatrice.raby@univ-reims.fr

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Abstract

In many cancers, including lung carcinomas, Fragile histidine triad (Fhit) is frequently decreased or lost. Fhit status has recently been shown to be associated with elevated \textit{in vitro} and \textit{in vivo} invasiveness in lung cancer. Tumor cell invasion is facilitated by epithelial-mesenchymal transition (EMT), a process by which tumor cells lose their epithelial features to acquire a mesenchymal cell-like phenotype. In this study, the mechanism underlying Fhit-regulated EMT was deciphered. Using Slug knockdown, pharmacological inhibitors PD98059, PP1, and gefitinib as well as an anti-EGFR antibody, it was demonstrated that Fhit silencing in bronchial cells induced overexpression of two primary EMT-associated targets, MMP-9 and vimentin, to regulate cell invasion dependent on an EGFR/Src/ERK/Slug signaling pathway. Moreover, ectopic expression of Fhit in Fhit-deficient lung cancer cells down-regulated this pathway. Finally, an inverse correlation was observed between Fhit and phospho-EGFR levels in a cohort of human squamous-cell lung carcinoma specimens. These results demonstrate a Fhit-dependent mechanism in the control of EMT-regulated EGFR signaling.

\textbf{Implications:} This study adds new insight into the regulatory mechanism of EMT, a process known to increase resistance to conventional and targeted therapies in lung cancer.
Introduction

Lung cancer is the leading cause of death worldwide. The poor survival of lung cancer patients is mostly attributed to early metastasis and resistance to both conventional and targeted therapies. The metastatic progression of epithelial tumors is a complex process requiring tumor cell plasticity. In particular, the step of tumor cell invasion into surrounding stromal tissue is characterized by a dedifferentiation of tumor cells, largely known to involve epithelial-mesenchymal transitions (EMT). Tumor cells undergoing EMTs lose epithelial features and engage in mesenchymal cell mimicry similarly as observed during embryonic development (1-3). Indeed, during tumor invasion, cell-cell adhesion complexes, especially adherens and tight junctions, are frequently reorganized in tumor cells. On the other hand, the cytoskeleton is also affected and a de novo expression of the intermediate filament vimentin is frequently observed during EMT (4-6). Tumor cells also acquire degradative properties against extracellular matrix components through their ability to produce themselves high levels of proteases such as matrix metalloproteases (MMPs) among which MMP-2, MMP-9 and MMP-14 (7, 8). Specific transcription factors, such as those of the Snail, the Twist or the ZEB family, have now been largely involved in the regulation of EMT target genes (9). Despite the difficulty to prove the occurrence of EMT in vivo because of its transient nature, several studies have reported changes of expression of EMT markers in non-small cell lung carcinomas (10, 11). There is growing evidence that EMT increases resistance to conventional chemotherapy, radiotherapy as well as targeted therapy (6). Accordingly, it has been shown that tumor cells of lung carcinomas which have undergone an EMT are less sensitive to epidermal growth factor receptor (EGFR) tyrosine kinase inhibitors (TKIs), gefitinib and erlotinib, than epithelial tumor cells (10, 12, 13).

The Fragile histidine triad (Fhit) gene is one of the most frequently altered genes in cancers, in particular in lung carcinomas in which Fhit promoter methylation may be a
prognostic marker (14, 15). The role of Fhit as a tumor suppressor gene has been well documented. Indeed, restoration of Fhit suppresses tumorigenicity in tumor cell lines and in mouse models through induction of caspase-dependent apoptosis and inhibition of proliferation of tumor cells (16-18). Besides its oncosuppressor activity, Fhit plays also a role in metastatic progression. Indeed, we previously showed that Fhit downregulation in non-small cell lung carcinomas is correlated with tumor invasion according to the tumor-node-metastasis (TNM) stage, particularly the lymph node status. Moreover, we demonstrated that Fhit controls invasion of lung tumor cells by regulating the expression of genes associated with EMT including cell-cell adhesion molecules, MMPs and vimentin (19). However, the mechanisms underlying this regulation process remains unknown.

In the present study, we intended to decipher the signaling pathway engaged by Fhit to regulate lung tumor invasion. We here showed that Fhit regulates two of its main targets, MMP-9 and vimentin, in bronchial cells along with cell invasion through an EGFR/Src/ERK/Slug signaling pathway.

Materials and Methods

Cell culture

The human lung cell lines HBE4-E6/E7 and NCI-H1299 were obtained from the American Type Culture Collection (Rockville, MD). Both cell lines were tested and authenticated by short tandem repeat profiling at the DSMZ (Braunschweig, Germany). Cells used for experiments were passaged for fewer than 6 months after resuscitation. HBE4-E6/E7 cells were cultured in Keratinocyte-SFM (Gibco, Invitrogen, Carlsbad, CA) supplemented with 0.2 ng/ml EGF and 25 µg/ml BPE. The stable transfectants NCI-H1299-mock and NCI-H1299-Fhit, generated as previously described (19), were cultured in RPMI (Gibco) containing 10% FCS.
Tumor tissue samples

Frozen tumor pieces for total protein extraction were obtained from samples of the Tumor bank belonging to the Reims University Hospital Biological Resource Collection n° DC-2008-374. Surgically resected tumors were collected after obtaining informed consent from 22 patients with squamous-cell lung carcinomas of stages I (6 cases), II (7 cases) and III (9 cases) according to the 2009 TNM classification. This study has been approved by the Institutional Review Board of the Reims University Hospital.

Pharmacological inhibitors and neutralizing antibodies

Pharmacological inhibitors of ERK1/2 (PD98059 and U0126), Src (PP1), p38 (SB203580), JNK (SP6600125), ROCK (Y27632), PKA (KT5720), PI3K (wortmaninn, KY12420) and PLC (U73122) pathways, the PDGFR tyrosine kinase inhibitor VII, the RTK inhibitor genistein and the FGFR tyrosine kinase inhibitor (PD166866) were purchased from Calbiochem (Merck, Darmstadt, Germany) and the EGFR tyrosine kinase inhibitor gefitinib (Iressa) from Tocris Bioscience (Bristol, UK). The concentrations used were 50 µM for PD98059, 20 µM for genistein, 10 µM for PP1, U0126, SB203580, and FGFR I, 2 µM for SP600125, 1 µM for Y27632, KT5720 and U73122, 0.8 µM for gefitinib, 0.1 µM for PDGFR I and 0.01 µM for wortmaninn. Controls were treated with DMSO. For EGFR neutralization experiments, a mouse monoclonal antibody to EGFR (2 µg/ml) (clone 225, Calbiochem, Merck, Darmstadt, Germany) was used. Non-immune mouse IgG1 (clone 1E2.2, Millipore, Temecula, CA) served as negative control.
Transfection of small interfering RNA

Three Fhit specific sequences were selected in the coding sequence of Fhit (GeneBank accession number: NM_002012) to generate 21-nucleotide sense and 21-nucleotide antisense strands of the type (19N) TT (N, any nucleotide). The selected 19-nt sequences were as follow: Fhit si1, 5’-CAUCUCAUCAAGCCCUCUG-3’; Fhit si2, 5’-GGAAGGCUGGAGACUUUCA-3’ and Fhit si3 5’-GGAGGACUUUCCUGCCUCU-3’ (Eurogentec, Seraing, Belgium). Three corresponding scrambled duplexes which do not recognize any sequence in the human genome were used as controls. A total of 100 000 HBE4-E6/E7 cells were transfected with a mix of the three siRNA duplexes (20 nM) by calcium phosphate-precipitation method as previously described (20). Two Slug specific siRNA si1: 5’-GCUACCCAAUGGCCUCUCU-3’ and si2: 5’-UCUGGCUGCUGUAGCAC-3’ were also used for co-transfection experiments. After transfection, cells were 24 hours BPE / EGF-starved. Cells were then submitted to an invasion assay or cultured for 24 hours in fresh BPE / EGF-free medium. For inhibition experiments, the various pharmacological inhibitors or the EGFR neutralizing antibody were added to the cells at the adequate concentration. Supernatants and cells were then harvested for zymography and Western blot analyses.

Western blotting

Total proteins from cells or tumor samples were extracted in RIPA buffer containing complete protease inhibitor cocktail (Roche diagnostics GmbH, Mannheim, Germany) and phosphatase inhibitor cocktail set II (Calbiochem, Merck, Darmstadt, Germany). Proteins were separated on SDS-PAGE gels and transferred to a PVDF membrane (NEN, Boston, MA). The membranes were incubated with either a goat polyclonal antibody to Slug (1:1000) (G-18, Santa Cruz Biotechnology, Santa Cruz, CA), or rabbit polyclonal antibodies to Fhit
(1:250) (ZR44; Zymed, Invitrogen, Carlsbad, CA) and ERK (1:1000) (C-16, Santa Cruz Biotechnology), or rabbit monoclonal antibodies to vimentin (1:1000) (clone SP20, Epitomics, Burlingame, CA), EGFR (1:10,000) (clone E235, Millipore, Temecula, CA) and phosho-Src family (Tyr416) (1:1000) (clone 100F9, Cell Signaling Technology, Danvers, MA) or mouse monoclonal antibodies to vimentin (1:1000) (clone V9, Dako, Carpinteria, CA), phospho-ERK (1:1000) (E-4, Santa Cruz Biotechnology), v-Src (1:1000) (clone 327, Calbiochem) and phospho-EGFR (Tyr1173) (1:500) (clone 9H2, Millipore). Following steps were performed as previously described.(20) Subsequent detection of GAPDH (using a mouse antibody, 1:75000, clone 6C5, Chemicon, Millipore, Billerica, MA) was performed on the same membranes as a control.

**Modified Boyden chamber invasion assay**

The *in vitro* invasive properties of cells were assessed using a modified Boyden chamber assay. $10^5$ cells were placed in the upper compartment of the invasion chamber (BD BioCoat Matrigel Invasion Chamber, BD Biosciences, Bedford, MA). For inhibition experiments, the various pharmacological inhibitors or the EGFR neutralizing antibody were added to the cells in the upper compartment of the insert at the adequate concentration. The chambers were incubated for 24 hours at 37°C. The filters were then fixed in methanol and stained with hematoxylin. Quantification of the invasion assay was performed by counting the number of cells at the lower surface of the filters (23 fields at 400-fold magnification).

**Gelatin zymography analysis**

Supernatants of cells grown for 24 hours in BPE / EGF-free conditions were collected, centrifugated and separated on 10% polyacrylamide SDS gel containing 0.1% (w/v) gelatin. The gel was washed for 1 hour at room temperature in a 2% (v/v) TritonX-100 solution,
transferred to a 50mM Tris-HCl / 10 mM CaCl$_2$ (pH 7.6) buffer and incubated overnight at 37°C. The gel was stained in a 0.1% (w/v) Coomassie Blue (G250) / 45% (v/v) methanol / 10% (v/v) acetic acid solution and de-stained in a 10% (v/v) acetic acid / 20% (v/v) methanol solution.

**Immunoprecipitation and c-Src activity assay**

Transfected cells treated 1h with DMSO or 10 μM PP1 were lysed in immunoprecipitation lysis buffer (10 mM Tris pH 7.4, 150 mM NaCl, 5 mM EDTA, 10 % (v/v) glycerol, 1 % (v/v) Brij 98) containing complete protease inhibitor cocktail (Roche diagnostics GmbH) and phosphatase inhibitor cocktail set II (Calbiochem). 300 μg of total proteins were incubated with 2.5 μg of anti-c-Src rabbit monoclonal antibody (Cell Signaling Technology) for 1 h at 4°C. The antigen-antibody complexes were incubated with protein G-Plus-Sepharose (Santa Cruz Biotechnology) for 1h at 4°C and washed three times in immunoprecipitation lysis buffer and twice with kinase buffer (20 mM Hepes pH 7.2, 10 mM MgCl$_2$, 10 mM MnCl$_2$, 1 mM Na$_3$VO$_4$). Pellets were resuspended in 50 μl kinase buffer supplemented with 5 μg denatured enolase (c-Src substrate) (Sigma-Aldrich, St Louis, MO), 100 μM ATP and 10 μCi [$\gamma$-$^{32}$P]-ATP (Perkin-Elmer, Waltham, MA). The reaction was performed for 30 min at 30°C and stopped by adding Laemmli sample buffer. Samples were resolved on a 10% SDS-PAGE, gels were dried and bands were visualized by autoradiography.

**Vimentin promoter-reporter assay**

The 1.5-kb vimentin promoter firefly luciferase reporter vector has been characterized previously (21). Transient transfections in NCI-H1299-mock and NCI-H1299 cells and luciferase activity assay were performed as previously described (20). The firefly luciferase
activity was normalized to the activity of the Renilla luciferase used as internal control. The data are expressed as fold induction relative to the empty vector-transfected NCI-H1299-mock cells.

**Src transduction**

HBE4-E6/E7 cells were transduced with ready-to-use lentiviral particles expressing mouse Src (CMV-mSRC-RB, LVP350) or negative control lentiviral expression particles (CMV-Null-RB) at a MOI of 10 (Amsbio, Oxon, UK). Stable transfectants were generated using blasticidin at a concentration of 3 µg.ml\(^{-1}\).

**Statistical analysis**

For *in vitro* experiments, data are expressed as a mean of fold induction ± standard error of mean and the one-sample *t*-test was used for statistical analyses. *In vivo* association between Fhit and pEGFR was tested with the use of non-parametric Spearman’s rank-correlation analysis (analysis based on the ranking of the samples). Since Fhit and pEGFR levels were not normally distributed, they were logarithmically transformed for graphic representation. *P* < 0.05 was considered significant.

**Results**

**Cell invasion induced by Fhit inhibition is Slug-dependent**

Previously, we reported that Fhit inhibition is able to increase cell invasion by regulating expression of genes associated with EMT, in particular vimentin and MMP-9 (19). We thus here searched for a potential regulation of transcription factors known to modulate EMT. We found that Slug is upregulated concomitantly with vimentin and MMP-9 after Fhit silencing in bronchial HBE4-E6/E7 cells as shown by western blot and zymography analyses (Figure
The functional implication of Slug was demonstrated by using Slug siRNA. In a modified Boyden chamber invasion assay, cell invasion induced by Fhit silencing was reduced by 66% ($P = 0.001$) and 40% ($P < 0.01$) when cells were co-transfected with Slug siRNA 1 and 2, respectively (Figure 1b and supplementary figures 1a and 2a and b). The inhibition of cell invasion by Slug siRNA 1 and 2 was correlated with a lower increase of vimentin and MMP-9 expression. The induction of vimentin by Fhit siRNA was reduced by 62% ($P < 0.001$) and 48% ($P < 0.05$) in the presence of Slug siRNA 1 and 2 and that of MMP-9 by 61% ($P < 0.001$) and 51% ($P < 0.01$), respectively (Figure 1c and supplementary figures 1b and 3a and b).

**Cell invasion induced by Fhit inhibition requires ERK1/2 signaling pathway**

Next, aiming at identifying the signaling cascade acting upstream the transcription factor Slug, we performed a large screening of different pharmacological inhibitors of known intracellular signaling pathways in a modified Boyden chamber invasion assay (Supplementary figure 4). We found that Fhit siRNA-induced cell invasion was extracellular signal-regulated kinase (ERK)-dependent. Indeed, the invasive capacities of Fhit-silenced cells were decreased by 64% in the presence of the PD98059 inhibitor ($P < 0.05$) (Figure 2a and supplementary figure 2c). Another inhibitor of the same pathway, U0126, was also able to reduce by 74% cell invasion induced by Fhit siRNA transfection ($P < 0.05$) (data not shown). Moreover, the increase of ERK 1/2 phosphorylation by Fhit silencing was demonstrated by Western blot analysis (Figure 2b). The effect of PD98059 was also observed on the expression level of Fhit targets. Indeed, a 65% reduction of vimentin level ($P < 0.01$) was observed in Fhit-silenced cells treated with PD98059 and a 54% for MMP-9 ($P < 0.05$). The level of Slug was also affected by the PD98059 treatment (reduction by 41%, $P < 0.05$)
supporting a cascade of regulation in which ERK acts upstream to Slug (Figure 2b and supplementary figure 3c).

Cell invasion induced by Fhit inhibition depends on Src

In addition, we observed that PP1, a Src kinase inhibitor, was as potent as PD98059 to decrease cell invasion in Fhit-silenced cells (by 66%, \( P < 0.01 \)) (Figure 3a and supplementary figure 2d) in comparison with other pharmacological inhibitors (Supplementary figure 4). Src activation by Fhit silencing was confirmed by the increase of Src phosphorylation as demonstrated by Western blot analysis (Figure 3b). PP1 inhibitor effect on cell invasion was correlated with a downregulation of Fhit targets. The induction of vimentin, MMP-9 and Slug expression by Fhit inhibition was reduced by 77% (\( P < 0.05 \)), 63% (\( P < 0.01 \)) and 65% (\( P < 0.05 \)), respectively (Figure 3b and supplementary figure 3d). Moreover, the fact that the phosphorylation state of ERK 1/2 is affected by PP1 suggests that Src acts upstream ERK 1/2 in the signaling cascade (Figure 3b). Combined treatment with both inhibitors, PD98059 and PP1, had no significant additional effect on cell invasion thus supporting this hypothesis (data not shown). Because PP1 and phospho-Src antibody target several Src family tyrosine kinase members, the specificity of Src involvement was confirmed by an in vitro kinase assay using enolase as Src substrate on e-Src immunoprecipitated from lysates of scrambled and Fhit siRNA-transfected cells (Figure 3c).

Cell invasion induced by Fhit inhibition involves EGFR

Finally, we investigated which cell surface receptor is implicated in activation of this Src/ERK/Slug cascade. We found that the increase of invasive capacities induced by Fhit silencing in bronchial cells HBE4-E6/E7 is 59% reduced by genistein thus suggesting the implication of receptor tyrosine kinase(s) (RTK)s (\( P < 0.05 \)) (Figure 4a). Indeed, the selective
inhibitor of EGFR tyrosine kinase, gefitinib, decreases by 51% invasion of Fhit-silenced cells \( (P < 0.01) \) whereas other RTK inhibitors, fibroblast growth factor receptor (FGFR) and (platelet-derived growth factor receptor (PDGFR) inhibitors, have no significant impact (Figure 4a and supplementary figure 2e). Moreover, the increase of EGFR phosphorylation by Fhit silencing was demonstrated by Western blot analysis (Figure 4b). Upregulation of vimentin, MMP-9 and Slug after Fhit siRNA transfection is respectively 35\% \( (P < 0.05) \), 48\% \( (P < 0.001) \) and 50\% \( (P < 0.01) \) lower in the presence of gefitinib (Figure 4b and supplementary figure 3e). We also observed that gefitinib influences the phosphorylation state of Src and ERK kinases, supporting a link with the Src/ERK cascade depicted above (Figure 4b). In addition, we found that the effect of EGFR inhibition by gefitinib on vimentin and MMP-9 expression is counteracted by Src overexpression in Fhit siRNA-transfected cells (Supplementary figure 5). Results obtained with gefitinib on cell invasion and Fhit targets were reproduced with an EGFR neutralizing antibody, thus precisely involving EGFR in the signaling pathway (Figure 4a and c and supplementary figures 2f and 3f). Furthermore, the ectopic expression of Fhit in Fhit-deficient lung cancer cells NCI-H1299 previously described to decrease their invasive capacities (19), induces a downregulation of the EGFR/Src/ERK/Slug signaling pathway concomitantly with a decrease of MMP-9 production and vimentin promoter activity (Figure 5). Finally, the \textit{in vivo} relevance of these findings was assessed by demonstrating by Western blot analysis an inverse correlation between Fhit and phospho-EGFR levels in protein extracts from a series of squamous-cell lung carcinoma samples \( (r = -0.4478, P < 0.05) \), especially those of TNM stage III \( (r = -0.8000, P < 0.05) \) (Figure 6).
Discussion

In this study, we demonstrated that one of the pathways engaged by Fhit to control lung tumor invasion is the regulation of two of its main target genes, MMP-9 and vimentin, by an EGFR/Src/ERK/Slug signaling cascade (Figure 7).

First, we found that Fhit silencing induces MMP-9 and vimentin overexpression along with cell invasion in a Slug-dependent way. Slug is a member of the Snail family of transcription factors known to be key mediators of EMT during both physiological processes such as embryonic development and pathological processes such as metastatic progression (22, 23). Slug has been particularly well described in the context of lung cancer progression. Indeed, Slug overexpression is an indicator of poor overall survival in lung cancer patients and, in agreement with our observations, Slug promotes invasion of lung cancer cells in vitro and in mouse models (13). Moreover, it was shown that Slug facilitates invasion of both pancreatic and oral cancer cells through upregulation of MMP-9 (24, 25). In addition, Vuoriluoto et al. previously demonstrated that Slug induces vimentin expression and migration in mammary cells (26). Vimentin, cell invasion and migration were also found regulated by exogenous modulation of Slug in a leiomyosarcoma cell line (27).

Also, we demonstrated that ERK signaling acts upstream Slug in this cascade of regulation. Accordingly, Slug expression is controlled by Erk5 in keratinocytes during wound healing (28). A direct link between ERK activation and Slug induction was also shown to play a role in E-cadherin repression in colon adenocarcinoma cells (29). Chen et al. showed that ERK pathway regulates breast cancer cell migration by maintaining Slug expression (30). A ERK/Slug pathway is involved in a mesenchymal-to-epithelial transition induced by clusterin silencing in lung adenocarcinoma cells (31). Finally, an association between ERK1/2 signaling and MMP-9 regulation by Slug was demonstrated by Joseph et al. (25).
Furthermore, we showed a link between Src and ERK signaling in the regulation of MMP-9, vimentin and cell invasion. This finding was supported by previous studies demonstrating a c-Src-mediated migration/invasion of breast cancer cells and hepatocellular carcinoma cells by downstream activation of the ERK pathway (32, 33). Src and MEK inhibitors have a combined effect on melanoma invasion (34). Src is overexpressed or hyperactivated in cancers of which lung carcinomas and is tightly associated with neoplastic invasion and EMT (35). Indeed, Src is known to promote the expression of MMPs such as MMP-9 by activating ERK and PI3K signaling (36). Src modulation also impacts vimentin expression in rat squamous bladder carcinoma cells and Wei et al. described a cross-linking between vimentin and c-Src in mediating prostate cancer invasion and metastasis (37, 38).

Finally, we identified EGFR as the receptor orchestrating the Src/ERK/Slug pathway modulated by Fhit silencing and/or Fhit ectopic expression. Altogether, these results thus show that Fhit regulates EGFR signaling pathway thereby inhibiting tumor cell invasion through the modulation of MMP-9 and vimentin levels. This is in line with our in vivo data showing an inverse correlation between Fhit and phospho-EGFR level in squamous-cell lung carcinomas and the previous work of Arnoux et al. demonstrating the control of cell motility in keratinocytes by a EGFR/Erk5/Slug cascade (28). EGFR is considered as a major inducer of EMT (39). In that way, it has been shown that EGF acts in synergy with TGF-β1 to increase migration of human renal cells by upregulating MMP-9 through ERK signaling (40). In addition, EGFR activation promotes migration and invasion of head and neck squamous cell carcinoma cells by inducing vimentin and MMP-9 in a ERK1/2-dependent way (41). The mechanism by which Fhit regulates EGFR signaling remains unknown. Fhit and Src have been shown to reciprocally coimmunorecipitate (42). One hypothesis may therefore be that Fhit sequestrates Src, thereby impeding it to associate with and mediating the phosphorylation and hyperactivation of EGFR. Accordingly, Src, known to be a key downstream of activated
EGFR family members, is also able to form a complex with EGFR to potentiate its oncogenic activity (43). Interestingly, Fhit has also been described as a physiological target of Src upon stimulation of EGFR leading to its proteasome degradation (42, 44). Therefore, Fhit is a substrate of EGFR/Src and its downregulation by various mechanisms such as promoter methylation or loss of heterozygosity may also influence the EGFR/Src signaling thus creating a regulatory loop between Fhit, EGFR and Src to sustain invasion of carcinoma cells.

In conclusion, this study provides new insights into the role of Fhit in the control of invasive phenotype of lung tumor cells. Interestingly, our results show the involvement of EGFR in the Fhit-driven process of EMT regulation. A better understanding of the mechanisms of regulation of EGFR signaling may be particularly helpful in the context of lung cancer and EMT-associated resistance to targeted therapy.

**Acknowledgments**

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References


Figure legends

**Figure 1:** Cell invasion induced by Fhit inhibition is Slug-dependent. (a) Western blot analysis of Fhit, Slug, vimentin and GAPDH levels in HBE4-E6/E7 cells transfected with scrambled siRNA or Fhit siRNA. Zymography analysis of MMP-9 level in conditioned media of HBE4-E6/E7 transfectants. (b) Analysis of the effect of Slug siRNA 1 or a scrambled siRNA on the invasive capacities of HBE4-E6/E7 transfectants in a modified Boyden chamber invasion assay. (c) Western blot analysis of the effect of Slug siRNA 1 or a scrambled siRNA on Fhit, Slug, and vimentin levels in HBE4-E6/E7 transfectants. MMP-9 has been analyzed by zymography.

**Figure 2:** Cell invasion induced by Fhit inhibition requires ERK1/2 signaling pathway. (a) Comparison of the invasive properties of HBE4-E6/E7 cells transfected with scrambled siRNA or Fhit siRNA in the presence of DMSO or PD98059 inhibitor (50 µM) using a modified Boyden chamber assay. (b) Western blot analysis of Fhit, phospho-ERK 1/2, total ERK 1/2, Slug, vimentin levels and zymography analysis of MMP-9 levels in HBE4-E6/E7 transfected cells treated with DMSO or PD98059.

**Figure 3:** Cell invasion induced by Fhit inhibition depends on Src. (a) Comparison of the invasive properties of HBE4-E6/E7 cells transfected with scrambled siRNA or Fhit siRNA in the presence of DMSO or PP1 inhibitor (10 µM) using a modified Boyden chamber assay. (b) Western blot analysis of Fhit, phospho-Src, total Src, phospho-ERK 1/2, total ERK 1/2, Slug, vimentin levels and zymography analysis of MMP-9 levels in HBE4-E6/E7 transfected cells treated with DMSO or PP1. (c) Activity of c-Src immunoprecipitated from lysates of HBE4-
E6/E7 transfected cells treated with DMSO or PP1 was assayed using enolase as substrate. Bands were visualized by autoradiography.

**Figure 4:** Cell invasion induced by Fhit inhibition involves EGFR. (a) Comparison of the invasive properties of HBE4-E6/E7 cells transfected with scrambled siRNA or Fhit siRNA in the presence of DMSO or either genistein (10 µM), a PDGFR inhibitor (0.1 µM), a FGFR inhibitor (2 µM) or gefitinib (0.8 µM) or in the presence of IgG1 or a neutralizing EGFR antibody (2 µg/ml) using a modified Boyden chamber assay. (b) Western blot analysis of Fhit, phospho-EGFR, total EGFR, phospho-Src, total Src, phospho-ERK 1/2, total ERK 1/2, Slug, vimentin levels and zymography analysis of MMP-9 levels in HBE4-E6/E7 transfected cells treated with DMSO or gefitinib. (c) Western blot analysis of Fhit, phospho-EGFR, total EGFR, phospho-Src, total Src, phospho-ERK 1/2, total ERK 1/2, Slug, vimentin levels and zymography analysis of MMP-9 levels in HBE4-E6/E7 transfected cells treated with IgG1 or a neutralizing EGFR antibody.

**Figure 5:** Ectopic expression of Fhit in NCI-H1299 cells induces a downregulation of the EGFR/Src/ERK/Slug signaling pathway. (a) Western blot analysis of Fhit, phospho-EGFR, total EGFR, phospho-Src, total Src, phospho-ERK 1/2, total ERK 1/2, Slug and GAPDH levels in NCI-H1299-mock and NCI-H1299-Fhit stable transfectants. (b) Zymography analysis of MMP-9 levels in NCI-H1299-mock and NCI-H1299-Fhit stable transfectants. (c) Vimentin promoter reporter assay. NCI-H1299-mock and NCI-H1299-Fhit transfectants were subjected to a transient transfection with a 1.5-kb vimentin promoter firefly luciferase reporter vector followed by a luciferase activity assay. The firefly luciferase activity was normalized to the activity of the Renilla luciferase used as internal control. The data are expressed as fold induction relative to the empty vector-transfected NCI-H1299-mock cells. *P < 0.05
Figure 6: Inverse correlation between Fhit and pEGFR levels in squamous-cell lung carcinomas. (a) Fhit and phospho-EGFR levels were analyzed by western blotting in protein extracts from a series of 22 SCLC human samples (6 TNM stage I, 7 stage II and 9 stage III). (b) The correlation coefficient $r$ was calculated using a Spearman’s rank-correlation analysis in the entire tumor series and also in the TNM stage III tumors.

Figure 7: Proposed mechanism of Fhit-regulated EMT process. Fhit silencing induces EMT-associated cell invasion by upregulating vimentin and MMP-9 via EGFR transactivation through the cytoplasmic effectors Src and ERK1/2 and the transcription factor Slug.
Joannes et al., Figure 1
Joannes et al., Figure 2

(a) Cell invasion

<table>
<thead>
<tr>
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<th>Scrambled</th>
<th>Fhit siRNA</th>
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<tr>
<td>DMSO</td>
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<td>PD98059</td>
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(b) Western blots

- Fhit
- pERK 1/2
- tERK 1/2
- Slug
- Vimentin
- GAPDH
- MMP-9
**Joannes et al., Figure 3**

**a**

Cell invasion

Fold induction

- Scrambled
- Fhit siRNA

DMSO  PP1

**b**

DMSO  PP1

Scrambled  Fhit siRNA  Scrambled  Fhit siRNA

- Fhit
- pSrc
- tSrc
- pERK 1/2
- tERK 1/2
- Slug
- Vimentin
- GAPDH
- MMP-9

32P-Enolase

Vimentin

Slug

GAPDH

MMP-9
Joannes et al., Figure 4
Joannes et al., Figure 5
a

TNM stage:

I          II          III

Fhit

pEGFR

b

TNM stages I + II + III

r = -0.4478
P = 0.0366

TNM stage III

r = -0.8000
P = 0.0138

Joannes et al., Figure 6
Joannes et al., Figure 7
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