Distinct Effects of Adipose-Derived Stem Cells and Adipocytes on Normal and Cancer Cell Hierarchy

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Abstract

Adipose-derived stem cells (ASC) have received considerable attention in oncology because of the known direct link between obesity and cancer as well as the use of ASCs in reconstructive surgery after tumor ablation. Previous studies have documented how cancer cells commandeर ASCs to support their survival by altering extracellular matrix composition and stiffness, migration, and metastasis. This study focused on delineating the effects of ASCs and adipocytes on the self-renewal of stem/progenitor cells and hierarchy of breast epithelial cells. The immortalized breast epithelial cell line MCF10A, ductal carcinoma in situ (DCIS) cell lines MCF10DCIS.com and SUM225, and MCF10A-overexpressing SRC oncogene were examined using a mammosphere assay and flow cytometry for the effects of ASCs on their self-renewal and stem-luminal progenitor-differentiated cell surface marker profiles. Interestingly, ASCs promoted the self-renewal of all cell types except SUM225. ASC coculture or treatment with ASC conditioned media altered the number of CD49fhigh/EpCAMlow basal/stem-like and CD49fmedium/EpCAMmedium luminal progenitor cells. Among multiple factors secreted by ASCs, IFNγ and hepatocyte growth factor (HGF) displayed unique actions on epithelial cell hierarchy. IFNγ increased stem/progenitor-like cells while simultaneously reducing the size of mammospheres, whereas HGF increased the size of mammospheres with an accompanying increase in luminal progenitor cells. ASCs expressed higher levels of HGF, whereas adipocytes expressed higher levels of IFNγ. As luminal progenitor cells are believed to be prone for transformation, IFNγ and HGF expression status of ASCs may influence susceptibility for developing breast cancer as well as on outcomes of autologous fat transplantation on residual/dormant tumor cells.

Implications: This study suggests that the ratio of ASCs to adipocytes influences cancer cell hierarchy, which may impact incidence and progression. Mol Cancer Res; 14(7); 660–71. ©2016 AACR.

Introduction

Adipose stem cells (ASC) are increasingly being used for aesthetic and reconstructive surgery in multiple surgical disciplines (1–3). Their main utility lies in the repair of soft-tissue contour deficits either as a result of aging (aesthetic) or tumor removal, trauma, and congenital malformations (reconstructive). While their use in cosmetic surgery is less controversial, there are some lingering doubts about their safety when used at sites of tumor extirpation. In addition, injection of fat into hormonally sensitive sites such as the breast could carry long-term consequences. Long-term follow-up data in these patients are still not available to ascertain the safety of ASCs in these patients. Nonetheless, it is important to investigate the effects of ASCs and adipocytes on cancer cells because of the established link between obesity and cancer (4). A recent review in Cancer Research identified 37 articles that investigated the influence of ASCs on cancer progression and metastases (5). ASC-derived factors such as VEGF, IL6, IL8, TNFα, hepatocyte growth factor (HGF), leptin, and insulin-like growth factor (IGF) are suggested to promote angiogenesis, inflammation, cell proliferation, and hypoxia in the tumor microenvironment (5–7). Reciprocally, breast tumor cells promote transdifferentiation of ASCs into cancer-associated fibroblasts, which enhance tissue stiffness (8, 9).

In contrast to ASCs, the role of adipocytes in cancer progression remains controversial. Many reports indicated protumor growth effect including adipocyte-mediated induction of epithelial-to-mesenchymal transition (EMT) and resistance to ERBB2-targeted therapy (10, 11). The adipokine lipocalin2 secreted by adipocytes linked obesity to breast cancer progression (12). However, a recent study showed reduced proliferation of breast cancer cells that are in direct contact with neighboring adipocytes (13).

There are several gaps in our understanding of the role of ASCs and adipocytes on breast cancer. The first, do normal and tumor cells respond similarly to ASCs? Second, to what extent do ASCs alter stem-luminal progenitor-differentiated cell hierarchy in normal and cancer breast epithelial cells? Addressing this question is critical because most breast cancers are believed to originate from luminal progenitor cells (14, 15). Third, do ASCs and adipocytes have distinct effects on normal and tumor cells and if so, can these
To address the above questions, we utilized an in vitro system where we measured the effects of conditioned medium (CM) from ASCs and ASC-derived adipocytes on stem-luminal progenitor-differentiated cell hierarchy under growth conditions that promote self-renewal as well as differentiation of stem cells. Spontaneously immortalized breast epithelial cell line MCF10A, MCF10A cells with enhanced EMT characteristics due to overexpression of SLUG (MCF10A-SLUG; ref. 16), a MCF10A derivative that generates ductal carcinoma in situ (DCIS: MCF10DCIS. com; ref. 17) and MCF10A cells transformed by the SRC oncogene (MCF10A-SRC) were tested (18). We demonstrate that ASCs and adipocytes have distinct effects on the hierarchy of immortalized and transformed breast epithelial cells. HGF, secreted at higher levels by ASCs, and IFNγ, secreted at higher levels by adipocytes, altered breast stem-luminal progenitor-differentiated cell profiles. We propose that the interindividual differences in the ratio of ASCs to adipocytes in fat tissue or the ability of ASCs to differentiate into adipocytes could influence the rate of differentiation of breast epithelial cells with ultimate impact on breast cancer incidence and progression.

Materials and Methods

Cell lines and mammosphere assay
MCF10A and MCF10DCIS.com cell lines were purchased from ATCC. MCF10A cells—overexpressing SLUG have been described previously (16). Dr. Kevin Struhl (Harvard Medical School; Boston, MA) provided MCF10A cells—overexpressing SRC oncogene linked to the ligand-binding domain of estrogen receptor (MCF10A-SRC) and these cells have been described previously (18). SUIM225 cells were purchased from Asterand and maintained in media recommended by Asterand. Primary ASCs were isolated from abdomen, hips, axilla, and/or flank and cultured as described previously (19). SUIM225 and MCF10DCIS.com cells were purchased for this project alone and were cultured for the first time for this study. MCF10A cells have been authenticated within the last 2 years using STR Systems for Cell line identification (DNA Diagnosis Center). Supplementary Table S1 provides details of sites of fat tissue collection, age, and BMI of individuals from whom ASCs were collected. All procedures of collecting human adipose tissue were approved by the Indiana University Human Research Review Board. All samples were from females. These ASCs have been characterized by flow cytometry for cell surface markers and for osteogenesis and adipogenesis as described previously (19, 20). ASCs were subjected to adipocyte differentiation by culturing in adipogenic differentiation media (21). CM from confluent cultures was used. The mammosphere assay has been described previously (22). Briefly, 5,000 cells were plated on ultralow adherent 6-well plates in MammoCult media from Stemcell Technologies. In coculture experiments, equal number of ASCs and GFP-labeled MCF10A or MCF10A-SRC cells were plated. Mammospheres were imaged and/or subjected to flow cytometry 6 to 10 days after plating.

Cytokines/chemokines/growth factors, antibodies, and flow cytometry

Human cytokine array panel A (R&D Systems, catalog no. ARY005) was used to measure secreted factors in CMs of ASCs and adipocytes. Assays were done as per instructions from the manufacturer. HGF, IFNγ, CXCL1, SERPINE1, IL17E, IL23, and CD40L were purchased from R&D Systems or Peprotech, Inc. Concentrations of cytokines/chemokines/growth factors used in mammosphere assays are indicated in figure legends. Antibodies against CD49f, CD44, CD24, and EpCAM used for flow cytometry have been described in our recent publication (23). Mammospheres were collected, washed, and exposed briefly to trypsin to obtain single-cell suspensions. Cells were stained with indicated antibodies and subjected to flow cytometry. The percentage of cells in each quadrant or region is presented in the article. For two-dimensional (2D) cultures, cells were treated with indicated cytokines/chemokines/growth factors for 48 hours, collected by trypsinization, and subjected to flow cytometry.

RNA isolation and qRT-PCR
RNA from isolated adipose tissue of individual donors was extracted using RNeasy kit (Qiagen). cDNA was synthesized using iScript cDNA synthesis kit (Bio-Rad). qRT-PCR was performed using TaqMan probes from Applied Biosystems. Primers used for qRT-PCR are as follows:

HGF: forward 5-ATT TCC CGT GTA G-3, reverse 5-CCC ATG GGT TGT -3.

IFNγ: forward 5-TGG GAA CCA GAT GCA AGT AAG-3, reverse 5-CAG GGT TGG GTT TTG ATT T-3.

CXCL1: forward 5-AAA CCG GCA GTA ACT GGA TAG-3, reverse 5-GTT GGT AGT AAC CAT GAG AAG TAT GA-3.

Statistical analysis

All data presented are mean ± SD from two to six experiments. In experiments with similar trends but with experimental variability, experiments with least variability were selected for statistical analysis. Student t test was used to measure the statistical significance between measurements (Graphpad.com).

Results

ASCs enhance self-renewal of breast epithelial cells

The mammosphere assay is used routinely as a surrogate assay to measure self-renewal; previous studies in neurosphere cultures have shown that the size of the spheres indicates the extent of self-renewal (24, 25). We used this assay to determine the effect of ASCs on self-renewal of immortalized and transformed breast epithelial cells. As breast epithelial cells are marked with GFP, spheres formed by breast epithelial cells and ASCs could be easily distinguished by fluorescent microscopy. MCF10A cells formed smaller spheres and the size of the spheres were much larger when cocultured with ASCs (Fig. 1A). ASCs from three different individuals gave similar results. ASCs did not have consistent effects on number of spheres but the sphere size increased. For example, ASC #4 increased the size of spheres from 73 × 40 μm to 111 × 80 μm. Furthermore, ASCs increased the size of the MCF10A-SRC–derived mammospheres (from 80 × 69 μm to 170 × 144 μm). We next determined whether CM from ASCs similarly influenced the size of mammospheres. Indeed, CM treatment alone was sufficient to increase the size of mammospheres (Fig. 1B). Thus, transacting soluble factors from ASCs are sufficient to enhance the self-renewal of immortalized and transformed breast epithelial cells.

ASCs expand basal/luminal progenitor cell population of MCF10A cells

Breast epithelial cells with basal/stem-like, luminal progenitor, and nonlonogenic/differentiated characteristics display CD49f+/EpCAM−, CD49f+/EpCAM+, and CD49f−/EpCAM+ cell surface marker profile, respectively (26, 27). We used these two
marker combinations to determine the effects of ASCs on phenotype of cells in the mammospheres. ASCs themselves do not stain for CD49f or EpCAM (data not shown). Nonetheless, we examined the phenotype of all cells in the coculture or cells gated for GFP positivity. ASC coculture increased the number of CD49f<sup>+</sup>/EpCAM<sup>+</sup> MCF10A cells (Fig. 2A). Results of two independent coculture experiments are shown in Fig. 2B. ASC CM also increased the number of CD49f<sup>+</sup>/EpCAM<sup>+</sup> cells (Fig. 2A).

We recently reported that the EMT-associated gene SLUG increases MCF10A stemness (16). As EMT is linked to various aspects of cancer progression including drug resistance (28, 29), we then examined whether ASCs can further alter the phenotype of MCF10A-SLUG cells. ASC CM increased the size of MCF10A-SLUG–derived mammospheres with corresponding elevation in number of cells with luminal progenitor phenotype (Fig. 2C).

**Transformed cells respond differently to ASC and adipocyte CMs**

To determine whether ASCs and adipocytes have different effects on breast epithelial cells, we obtained ASCs from three additional individuals and collected CMs from undifferentiated and adipocyte-differentiated ASCs. CMs from ASCs were more effective than CMs from adipocytes in increasing the size of MCF10A-derived mammospheres (Fig. 3A). ASC CM (all three) but not adipocyte CM (one of three) increased the number of CD49f<sup>+</sup>/EpCAM<sup>+</sup> cells in the mammospheres (Fig. 3B and C).

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**Figure 1.** ASCs increased the size of mammospheres. A, effects of ASC coculture on the size of mammospheres derived from MCF10A and MCF10A-SRC cells. Both cell lines express GFP and GFP-positive spheres were visualized under fluorescent microscope. ASCs themselves formed spheres, which were not fluorescent (bottom). B, ASC CM increased the size of mammospheres. CM from ASC #4 was used in this assay.
ASCs increased the number of MCF10A and MCF10A-SLUG cells with luminal progenitor features. A, CD49f/EpCAM staining pattern of GFP-expressing MCF10A cells with or without ASC #4 coculture or CM. Under both conditions, ASCs increased CD49f\(^{+}\)/EpCAM\(^{+}\) cells (with or without gating for GFP\(^{+}\) cells).

B, results of two independent experiments are shown.

C, ASC CM increased the size of spheres and the luminal progenitor pool of MCF10A-SLUG cells. Image 10\(\times\) in all figures. Results from three independent experiments are shown (right side).
Figure 3.
ASCs and adipocytes displayed distinct effects on MCF10A cells. A, effects of ASC and adipocyte CMs on MCF10A-derived mammospheres. Although MCF10A cells do not efficiently form the mammospheres, spheres formed in the presence of ASC CMs were larger in size. B, ASC CM increased the number of CD44+/CD24− and CD49f+/EpCAM− stem cells. Adipocyte CM was less efficient in inducing these changes. C, ASC CM was more efficient than adipocyte CM in increasing CD49f+/EpCAM− cells in mammospheres. D, ASC CM was more efficient than adipocyte CM in increasing the number of CD44+/CD24− cells in mammospheres. E, ASC and adipocyte CMs did not alter CD49f/EpCAM profile of MCF10A cells under 2D culture condition. Cells were maintained in DMEM/F12 with 10% FBS for 5 hours without any supplements, switched to serum-free media for 48 hours and then treated with CM for 48 hours. F, ASC and adipocyte CMs did not alter the CD44/CD24 profile of MCF10A cells under 2D culture condition.
Note that these three samples were obtained from individuals with higher BMI and ASCs from these individuals increased stem rather than luminal progenitor cells (Compare Figs. 2 and 3). Thus, there may be interindividual variation in the effect of ASCs rather than luminal progenitor cells (Compare Figs. 2 and 3).

ASCs increase the number of luminal progenitors in DCIS cells

The cell line SUM225 is also being used as a model for DCIS (6). HGF and IFNγ increased the number of CD49f/C0 cells (Fig. 5A and B). Differentiation into adipocytes reduced the size of mammospheres, whereas IFNγ reduced the size of mammospheres (Supplementary Fig. S4A). None of the other factors had an effect on the size of mammospheres (data not shown). In MCF10A cells, HGF had no effect on the size of the mammospheres, whereas IFNγ reduced the size of mammospheres (Supplementary Fig. S4A).

We next determined the phenotypic effect of HGF and IFNγ on cells in the mammosphere. Similar to the effects of ASC CM, HGF altered CD49f/C0/EpCAM+ and CD49f/C0/EpCAM− ratio in favor of elevated CD49f+/EpCAM+ progenitor MCF10DCIS.com cells (Fig. 6B and C). Similar effects were noted with IFNγ, although there was marked experimental variability. Note that neither HGF nor IFNγ altered CD49f/C0/CD24 staining pattern in MCF10DCIS.com-derived mammospheres, although overall CD49f staining intensity was higher in IFNγ-treated and lower in IFNγ-treated mammospheres.

HGF and IFNγ displayed distinct effects on the phenotype of MCF10A-SRC-derived mammospheres. While HGF increased CD49f+/EpCAM+ luminal progenitor cells, IFNγ increased the number of stem cells (Supplementary Fig. S3B). In fact, IFNγ increased the intensity of CD49f staining, whereas HGF increased EpCAM staining intensity (Supplementary Fig. S3C). In case of MCF10A cells, HGF increased the number of CD49f+/CD24− cells similar to ASC CM albeit weakly (Supplementary Fig. S4B). IFNγ increased the number of CD49f+/EpCAM+ progenitor and CD49f+/EpCAM− stem/basal cells (Supplementary Fig. S4C and D). It appears that IFNγ blocked differentiation of stem/progenitor cells as mammospheres had no CD49f+/EpCAM+ cells. These results show that HGF and IFNγ distinctly alter the stem or progenitor cell composition of immortalized, DCIS, and invasive cancer cells. As the effects of ASC CM and HGF/IFNγ overlapped partially, it appears that combined effects of multiple factors in ASC CM contribute to mammosphere expansion.

To determine whether there is any relationship between HGF and IFNγ expression in ASCs and adipocytes, we measured their mRNA levels by qRT-PCR. Differentiation of ASC to adipocytes caused significant drop in HGF levels with concomitant increase in IFNγ levels (Fig. 6D).

HGF/IFNγ mRNA ratio in the breast tumor microenvironment predicts outcome in patients with ER+ or PR− breast cancers

As adipocyte differentiation altered the ratio between HGF and IFNγ, we next determined whether the ratio between these two factors within the breast cancer environment has any prognostic
Figure 4. CM from both ASC and adipocytes increased the size of MCF10A-SRC-derived mammospheres. A, phase contrast images of mammospheres under three conditions (10×). B, CD49f/EpCAM and CD44/CD24 profiles of MCF10A-SRC-derived mammospheres under ASC or adipocyte CM-treated conditions. Both ASC and adipocyte CMs increased CD49f⁺/EpCAM⁺ cells. ASC and adipocyte CMs did not affect CD44 and CD24 status, as these spheres were predominantly CD44⁺/CD24⁻. Results using CM from ASC/adipocytes of patient #1 and 2 are shown. C, ASC and adipocyte CMs did not alter CD49f/EpCAM status of MCF10A-SRC cells grown under 2D condition. D, Adipocyte CM but not ASC CM increased CD44⁺/CD24⁻ MCF10A-SRC cells grown under 2D condition.
value using a tool we developed recently (34). HGF/IFNγ ratio can be a surrogate for ASCs and adipocytes composition in the tumor microenvironment. cBioportal analysis showed a tendency of mutual exclusivity in their expression in breast cancer (data not shown). In the TCGA dataset, elevated HGF/IFNγ ratio was associated with poor overall survival among patients with estrogen receptor (ER)-negative and progesterone receptor (PR)-negative but not ER-positive and PR-positive breast cancers (Supplementary Fig. S5). As ER-negative and PR-negative breast cancers are associated with poor outcome, ASC/adipocyte ratio may influence outcome in patients with these highly aggressive forms of breast cancer.

Figure 5.
CM from ASCs but not adipocytes increased the size of MCF10DCIS.com-derived mammospheres. A, phase contrast images of mammospheres under three conditions (10×). B, CD44/CD24 and CD49f/EpCAM staining pattern of cells from mammospheres. C, ASC CM increased CD49f−/EpCAM+ and CD44+/CD24− cells in patient-specific manner. Results of two or more experiments are shown. *, P < 0.05, control versus ASC CM-treated condition. D, adipocyte CM did not alter CD49f/EpCAM staining pattern of cells from mammospheres.
Discussion

Multiple mechanisms have been proposed to link adiposity with cancer incidence and/or outcome (4). Several protumorigenic and antitumorigenic factors have been identified in both ASCs and adipocytes. For example, adipocyte secreted leptin and adiponectin have opposing biologic effects in the normal breast, which impacts breast cancer susceptibility (35). Xenograft models have shown that the type of adipocytes, particularly beige/brown adipocytes, influence breast tumor development and progression (36). The majority of mechanistic studies on the effects of adiposity on breast cancer have focused on leptin signaling, including leptin-STAT3-G9a axis, leptin-induced EMT, and angiogenesis (37–39). Several other soluble factors from ASCs have been implicated in this process including VEGF, IL8, IGF, and HGF (5). A recent "meta PubMed" search identified 37 studies that have linked ASCs to cancer progression with the majority indicating unfavorable effects (5). Concurrently, breast cancer–derived factors themselves have been shown to alter plasticity and

Figure 6. HGF and IFNγ had distinct effects on MCF10DCIS.com mammospheres. A, effects of recombinant HGF (50 ng/mL) and IFNγ (50 ng/mL) on MCF10DCIS.com-derived mammospheres. Mammospheres were treated for 8 to 10 days with addition of new media & recombinant proteins every three days. B, CD44/CD24 and CD49f/EpCAM staining pattern of mammospheres with and without HGF and IFNγ treatment. C, both HGF and IFNγ increased the number of CD49f+/EpCAM+ cells. *, P < 0.007, control versus HGF or IFNγ; **, P < 0.05 HGF versus IFNγ. D, ASC to adipocyte differentiation caused reduction in HGF but an increase in IFNγ. All differences are statistically significant.
The Effect of Adipose Stem Cells on Breast Cancer

Adiposity has not only been linked to cancer incidence but also to the molecular subtypes of breast cancer (4). For example, in premenopausal women, adiposity is associated with approximately 10% risk reduction in less aggressive ER+/PR+ breast cancers but approximately 80% increase in risk of aggressive ER+/PR- and triple-negative breast cancers. In postmenopausal women, adiposity increases risk for both ER+/PR+ and ER-/PR- breast cancers. While multiple studies have independently analyzed the effects of ASCs and adipocytes, there has not been a comprehensive study comparing the effects of ASCs and adipocytes on “progression series” of breast epithelial cells, particularly in the context of epithelial cell hierarchy. Our study has addressed this issue by measuring the effects of ASCs and their differentiated counterparts on immortalized breast epithelial cells, noninvasive cancer cells representing DCIS, and invasive breast cancer cells. While ASCs were more effective than adipocytes in altering stem/progenitor cell composition of immortalized and noninvasive cancer cells, adipocytes were as effective as ASCs in altering progenitor/stem cell composition of invasive cancer cells. Please note that we did not observe a consistent increase in the number of mamsospheres by any treatment suggesting that ASCs do not cause dedifferentiation of differentiated cells to stem/progenitor cells. Instead, ASCs or adipocytes have an effect on the self-renewal or expansion of existing pool of stem/progenitor cells. We also observed a lack of effect of ASC or adipocytes on a DCIS cell line with ERBB2 amplification suggesting that certain cancer-associated genomic events activate signaling cascades that overlap with adipogenic-driven signals. Additional studies with multiple models are needed to identify the cancer subtypes that are not impacted by adiposity or ASC/adipocyte ratio.

The differential effect of ASC and adipocyte CM on noninvasive and invasive breast cancer cells is intriguing. We found ASC CM containing more soluble factors than adipocyte CM with very few being common to both. It is interesting that both ASC and adipocyte CM secrete IL23, a cytokine that suppresses immune response during tumor initiation, growth, and metastasis (41, 42). Therefore, ASCs and adipocytes may play a similar role in blocking host immune response to tumor. In addition, adiposity may have an impact on response to recently developed immune checkpoint therapy for cancer (43).

While the role of ASC-derived HGF on breast cancer cells has been studied (6), its influence on breast epithelial cell differentiation is unknown. Our studies indicate the ability of HGF to expand luminal progenitor cell population. Luminal progenitor cells in breast epithelial cell hierarchy are believed to be the most susceptible cell population for transformation and ASC composition in the breast may determine the proportion of cancer susceptible subpopulations of breast epithelial cells (44). Although HGF-mediated signaling events that promote expansion of luminal progenitor cells are yet to be elucidated, HGF-cMET axis has been shown to activate β-catenin, which is a critical transcription factor in breast stem/progenitor cells (6, 44).

We noticed unique effects of IFNγ on breast epithelial and cancer cells. Although we did not observe a uniform effect of IFNγ on the size of mamsospheres, it promoted the accumulation of stem-like or progenitor cells and possibly blocked differentiation of MCF10A cells. It is unclear at present whether IFNγ selectively kills differentiated cells, promotes dedifferentiation of differentiated cells, or blocks differentiation at progenitor cell state. Nonetheless, the observed effect of IFNγ is significant because its expression is induced upon differentiation of ASCs to adipocytes (Fig. 6) and its higher expression compared with HGF is associated with favorable outcome in ER- and PR- breast cancer patients (Supplementary Fig. S5). It is possible that IFNγ promotes dormancy of tumor cells by inhibiting their cell-cycle progression and maintaining them in G0-G1 stem-like state. IFNγ has previously been shown to inhibit cell-cycle progression of mammary epithelial cells by inducing the expression of p27kip1 (45). A recent study has shown stem cell-like properties of p27kip1-positive breast epithelial cells (46). Collectively, our studies suggest that ASC/adipocyte composition in the breast microenvironment determines HGF/IFNγ ratio, which ultimately has an impact on differentiation status of normal or cancerous breast epithelial cells. While this manuscript was being prepared for submission, Picon-Ruiz and colleagues showed that immature and mature adipocytes promote cytokine production and drive SRC/SOX2/miR302b-mediated stem cell signaling in breast cancer cells (47). However, the study did not investigate adipocyte-derived factors involved in promoting cytokine production by cancer cells. It is possible that HGF/IFNγ levels might determine to what extent ASCs/adipocytes influence feed-forward cytokine production loop in the breast environment.

Translation of these findings into clinical practice requires further study. While removal of the tumor is obviously important, reconstructive options to improve quality of life gain in meaning as long-term survival increases. One of the new options is the transplantation of adipose tissue augmented with or without ASCs into the breast extirpation site. This current study is the first to show that the ratio of adipocytes and ASCs may be a critical influence of breast cancer cell hierarchy, with some of the control over this decision in the hands of the reconstructive surgeon. However, individual variations in the ability of ASCs to differentiate to adipocytes, the levels of secreted HGF and IFNγ, as well as the genomic aberration status and/or molecular subtypes of any residual tumor cells may ultimately determine the safety of ASC/adipocyte transplantation. Characterization of ASCs and tumors on an individual basis may help to establish safety levels and provide rational guidelines for safer decisions.

Disclosure of Potential Conflicts of Interest
K. March has ownership interest (including patents) in S&K and NeuroFiX. No potential conflicts of interest were disclosed by the other authors.

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References

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