Heparin-like Polysaccharides Reduce Osteolytic Bone Destruction and Tumor Growth in a Mouse Model of Breast Cancer Bone Metastasis

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Abstract

TGF-β regulates several steps in cancer metastasis, including the establishment of bone metastatic lesions. TGF-β is released from bone during osteoclastic bone resorption and it stimulates breast cancer cells to produce osteolytic factors such as interleukin 11 (IL-11). We conducted a cell-based siRNA screen and identified heparan sulfate 6-sulfotransferase 2 (HS6ST2) as a critical gene for TGF-β–induced IL-11 production in highly bone metastatic MDA-MB-231(SA) breast cancer cells. HS6ST2 attaches sulfate groups to glucosamine residues in heparan sulfate glycosaminoglycans. We subsequently showed how heparin and a high-molecular-weight Escherichia coli K5–derived heparin-like polysaccharide (K5-NSOS) inhibited TGF-β–induced IL-11 production in MDA-MB-231(SA) cells. In addition, K5-NSOS inhibited bone resorption activity of human osteoclasts in vitro. We evaluated the therapeutic potential of K5-NSOS and fragmin in a mouse model of breast cancer bone metastasis. MDA-MB-231(SA) cells were inoculated into the left cardiac ventricle of athymic nude mice which were treated with fragmin, K5-NSOS, or vehicle once a day for four weeks. Both heparin-like glycosaminoglycans inhibited weight reduction, decreased osteolytic lesion area, and reduced tumor burden in bone. In conclusion, our data imply novel mechanisms involved in TGF-β induction and support the critical role of heparan sulfate glycosaminoglycans in cancer metastasis as well as indicate that K5-NSOS is a potential antimetastatic and antiresorptive agent for cancer therapy. This study illustrates the potential to translate in vitro siRNA screening results toward in vivo therapeutic concepts. Mol Cancer Res; 10(5); 597–604. ©2012 AACR.
targeted heparan sulfate 6-O-sulfotransferase 2 (HS6ST2), an enzyme that attaches sulfate groups to glucosamine residues in heparan sulfate. Therefore, we subsequently evaluated the effects of exogenous highly sulfated heparan sulfate glycosaminoglycans on TGF-β induction of IL-11. Highly sulfated heparan sulfate glycosaminoglycans, such as heparin, are commonly used to prevent venous thromboembolism, which is a common complication in patients with cancer as well. Heparin and heparin-like glycosaminoglycans (HLGAGs) have also been shown to prolong survival of patients with cancer, suggesting an antitumor effect for these compounds (3–5). The applicability of heparin in cancer therapy is limited because of its strong anticoagulant activity, but HLGAGs with less undesired effects on blood coagulation have been developed. Two HLGAGs with low anticoagulant activity, a low-molecular-weight (LMW) heparin fragmin and a high-molecular-weight (HMW) E. coli K5-derived heparin-like polysaccharide (K5-NSOS) were used in our studies. In addition to analyzing the effects of these HLGAGs on TGF-β–induced IL-11 production, we studied their in vivo antitumoral properties in a mouse model of breast cancer bone metastasis and effects on osteoclast activity in vitro.

**Materials and Methods**

**Cell culture**

MDA-MB-231(SA) cells were comprehensively characterized by comparative genomic hybridization and genomewide gene expression profiling by our research group as described in (6). The cells were cultured in Dulbecco’s Modified Eagle’s Medium supplemented with 10% inactivated calf serum, 1% penicillin/streptomycin, and nontoxic amino acids at +37°C in a humidified incubator in an atmosphere of 5% CO₂.

**siRNA transfection and IL-11 and cell viability assays**

Cells were transfected with siRNAs (Qiagen) at a final concentration of 15 (384-well plates) or 20 nmol/L (96-well plates). Briefly, the siRNAs were robotically printed to black 384-well clear-bottom plates or manually pipetted to black 96-well clear-bottom plates. siLenFect transfection agent (Bio-Rad Laboratories) diluted in OptiMEM (Gibco Invitrogen) was added into each well. After a 60-minute incubation at room temperature, 1,400 (384-well plates) or 10,000 (96-well plates) cells per well were added. After 24 hours, medium was changed and heparin, fragmin, or K5-NSOS (0.25 mg/mL) and TGF-β (5 ng/mL) were added. IL-11 concentration in the conditioned medium and cell viability were measured 24 hours later using the DuoSet ELISA Development System for human IL-11 (R&D Systems) and CellTiter-Blue assay (Promega), respectively, according to the manufacturer’s instructions.

**Quantitative reverse transcription RT-PCR**

RNA isolation, reverse transcription, and TaqMan analyses were done as previously described (6). The primers and probes are listed in the Supplementary Table S1.

**Smad luciferase reporter assays**

Smad signaling was quantified using the Cignal Smad Reporter Assay Kit (SA Biosciences). siRNAs (10 nmol/L) along with 93-ng luciferase reporter construct were cotransfected into MDA-MB-231(SA) cells using 0.47 μL Lipofectamine 2000 (Invitrogen). Medium was replaced with serum-free medium 18 hours after transfection, and 5 ng/mL TGF-β was added 23 to 27 hours later. Activity of the firefly luciferase reporter and Renilla luciferase was measured after 6- to 10-hour TGF-β induction using the Dual-Glo Luciferase Assay System (Promega Corporation) according to the manufacturer’s instructions. The background signal from the untransfected cells was subtracted, and the firefly reporter values were divided with the respective Renilla values.

**Heparin and HLGAGs**

Heparin from porcine intestinal mucosa was obtained from LEO Pharma and LMW heparin dalteparin (fragmin) from Pharmacia (ATC code B01A04, molecular weight 4–6 kDa). The HMW chemically N, O-sulfated bacterial polysaccharide (capsular polysaccharide from E. coli K5) was prepared as described in (7). The overall sulfation degree (sulfate groups per disaccharide unit) of the prepared K5-NSOS derivative was more than 3.5 and the molecular weight approximately 37 kDa.

**Mouse bone metastasis model**

Effects of fragmin and K5-NSOS on the development of bone metastases were studied using a previously described mouse model of osteolytic breast cancer bone metastasis (8, 9). Female nude mice (BALB/c nu/nu; Harlan) 4 to 5 weeks of age were inoculated with 100,000 MDA-MB-231(SA) cells into the left cardiac ventricle. Mice were administered with fragmin s.c. and K5-NSOS i.v. (5 mg/kg diluted in sterile water) or sterile water as a control (n = 11, 7, and 10 mice per group, respectively) at the time of cell inoculation and daily thereafter. The mice were maintained in a pathogen-free environment and monitored daily for clinical signs. Animal studies were conducted in accordance with the institutional animal care protocol or University of Texas Health Science Center at San Antonio (San Antonio, TX). Body weight was obtained at baseline and then weekly for 4 weeks, at which time the mice were sacrificed. Necropsy was conducted on all mice, and those with tumor in the chest were excluded from analysis because this indicated that the tumor inoculums did not properly enter the left cardiac ventricle. At sacrifice, hind and fore limbs and soft tissues were collected for histologic analysis.

**Radiographic analyses**

Development of bone metastases was monitored weekly by X-ray radiography. The animals were anaesthetized with Ketamine-Xylazine cocktail and X-rayed in a prone position with the Faxitron Specimen Radiographic System MX-20 DC-2 (Faxitron Corporation). The total lesion number and area in hind (left and right tibia and femur) and fore (left and right humerus) limbs per mouse were quantified from the images using MetaMorph image analysis software.
(Molecular Devices Corporation). All radiographs were evaluated without knowledge of treatment groups.

**Histology and histomorphometry**

The tissue samples were fixed in 10% neutral-buffered formalin for 2 to 3 days. The soft tissue samples were then stored at 70% ethanol, and the bone samples were decalcified in 14% EDTA for 2 weeks. After embedding in paraffin for 2 to 3 days, the soft tissue samples were then sectioned at 3.5 μm using a standard microtome. Tissue sections were placed on poly-L-lysine-coated glass slides and stained with hematoxylin and eosin (H&E) using standard histologic staining procedures. Some sections were stained with tartrate-resistant acid phosphatase (TRAP)-stained for determining the number of osteoclasts. Total tumor areas were measured from mid sagittal sections of tibiae, femora, and humerus. Total tumor area and total bone area per bone were determined using MetaMorph image analysis software. In addition, the number of osteoclasts per millimeter of tumor–bone interface in midsections of tibiae and femora was determined. Histomorphometry was conducted without knowledge of treatment groups.

**Osteoclast cultures**

Human osteoclast precursors (Poietics, Cambrex) were cultured on bovine bone slices (Nordic Bioscience Diagnostics) in 96-well plates for 10 days according to the manufacturer’s instructions. After the osteoclasts’ differentiation period, at day 7, the culture medium was replaced with fresh medium and different concentrations of fragmin or K5-NSOS were added. Osteoclasts were cultured for an additional 3 days. To determine the number of osteoclasts, TRACP 5b activity was measured from the medium collected at day 7 using the BoneTRAP Assay (IDS Ltd). At day 10, the amount of CTX (C-terminal telopeptide of α1 collagen) released from bone was quantified from the medium using CrossLaps for Culture assay (Nordic Bioscience Diagnostics). Resorption index was calculated by dividing the CTX values (day 10) by the TRACP 5b values (day 7; ref. 10). Multinuclear TRACP-positive osteoclasts were visualized by staining the nuclei with Hoechst 33258 (Sigma-Aldrich). Resorption pits were visualized by staining the nuclei with Hoechst 33258 (Sigma-Aldrich) and TRACP content of the cells using a leukocyte acid phosphatase kit (Sigma-Aldrich). Resorption pits were visualized using TRITC-conjugated WGA lectin (L-5266, Sigma), as previously explained (11). A cysteine protease inhibitor E64 (Sigma) was used as a reference inhibitor of osteoclast activity, as previously described (12).

**In vitro coagulation tests**

Anti-IIa and anti-Xa activities of fragmin, heparin, and K5-NSOS were determined using Chromogenic IL Test Heparin assay kit according to the manufacturer’s protocol and ACL 7000 automated coagulation analyzer (both from Instrumentation Laboratory Company). In the assays, the residual factors IIa and Xa were quantified with a synthetic chromogenic substrate. The buffer solution 0.9% NaCl was used in the anti-Xa activity measurements whereas buffer solution 0.05 mol/L Tris, 0.15 mol/L NaCl, 1% (w/v) bovine serum albumin, pH 7.4 was used in the anti-IIa measurements. The compounds were measured against an international LMW heparin standard ("Low-Molecular-Mass Heparin", European Pharmacopoeia Commission). The effects of fragmin, heparin, and K5-NSOS on activated partial thromboplastin time (APTT) in citrated plasma were measured using IL Test APTT Lyophilized silica kit (ILS Laboratories Scandinavia Oy) according to the manufacturer’s instructions. Dose–response curves as clotting time versus test compound concentration were prepared, and the doses causing a coagulation time of 100 seconds (APTT<sub>100s</sub>) were estimated from the trend lines.

**Statistical analysis**

The in vitro data are shown as mean ± SD and in vivo data as mean ± SEM. Data were analyzed for statistical significance by one-way ANOVA followed by the Student t test or the Dunn test or by nonparametric Kruskal–Wallis test followed by the Dunn comparison. P values less than 0.05 were considered statistically significant.

**Results**

**Identification of genes whose expression is critical to TGF-β–induced IL-11 production in MDA-MB-231 (SA) cells**

We noted an increased basal and TGF-β–induced IL-11 secretion in the highly bone metastatic MDA-MB-231(SA) variant as compared with the parental MDA-MB-231 breast cancer cell line (Supplementary Fig. S1). We used the genome-wide gene expression profiles of these cell types (6) and constructed a library consisting of 193 siRNAs targeting 107 genes (1 validated or 2 nonvalidated siRNAs per gene) that were most highly overexpressed in MDA-MB-231(SA) versus parental cells. In addition, we included a scrambled sequence siRNA as a negative control and 2 IL-11 siRNAs as positive controls. The most potent inhibitors of TGF-β–induced IL-11 secretion were confirmed using multiple replicates (Supplementary Table S2 and Fig. S2A). One such hit was an siRNA-targeting HS6ST2 (Fig. S1A and B). This gene encodes an enzyme that attaches sulfate groups to C-6 of glucosamine residues in heparan sulfate glycosaminoglycans. We confirmed the overexpression of HS6ST2 in the highly metastatic MDA-MB-231(SA) versus parental cells by quantitative RT-PCR (Supplementary Fig. S3A). We also studied whether other enzymes that attach sulfate groups to glycosaminoglycans in heparan sulfate glycosaminoglycans affect IL-11 secretion. Our data showed that knockdown of HS6ST1 had a similar effect as HS6ST2 silencing whereas knockdown of HS3ST1 slightly increased IL-11 secretion (Supplementary Fig. S2B). The knockdown efficiencies of the siRNAs were measured by quantitative RT-PCR (Supplementary Fig. S3B).

**HS6ST2 silencing inhibits Smad signaling**

Because TGF-β induction of IL-11 has been shown to be mediated by the Smad pathway (13, 14), we tested whether silencing of HS6ST2 affected Smad2/3/4-mediated transcriptional activity. This was done using a luciferase reporter...
construct containing a functional Smad-binding site. HS6ST2 silencing suppressed the TGF-β–induced luciferase signal, indicating a decrease in Smad binding to its response element (Fig. 1C).

Heparin and a HLGAG K5-NSOS inhibit TGF-β–induced IL-11 production in MDA-MB-231(SA) cells

It has been shown that TGF-β1 binds to heparin and highly sulfated liver heparan sulfate, and this potentiates the biologic activity of TGF-β1 (15–17). It is also well established that heparan sulfate–protein interactions critically depend on the amount and the positions of the sulfate groups (18). We therefore hypothesized that exogenous highly sulfated heparan sulfates, such as heparin and HLGAGs, may interfere with the physiologic heparan sulfate–TGF-β interactions and affect TGF-β signaling. Our in vitro data showed that heparin and a HMW HLGAG K5-NSOS inhibited TGF-β–induced IL-11 production in MDA-MB-231(SA) cells (P < 0.001) whereas a LMW HLGAG fragmin did not show a significant effect (Fig. 2).

K5-NSOS and fragmin inhibit weight reduction and reduce cachexia in tumor-bearing animals

We evaluated the therapeutic potential of K5-NSOS and fragmin in a mouse model of breast cancer bone metastasis. The mice that had been inoculated with MDA-MB-231(SA) cells were treated with fragmin, K5-NSOS, or vehicle at the time of MDA-MB-231(SA) cell inoculation and daily thereafter. The vehicle group had markedly lower body weight at sacrifice than in the baseline body weight due to tumor-induced cachexia. In contrast, the body weights of the fragmin- and K5-NSOS–treated mice at sacrifice were significantly higher than the vehicle group (P < 0.01 and P < 0.001; Fig. 3A). Furthermore, only 18% of the fragmin-treated mice and 29% of the K5-NSOS–treated mice were cachectic as compared with 86% of the vehicle-treated mice at sacrifice.

K5-NSOS and fragmin reduce osteolytic lesions and tumor growth in bone

Osteolytic lesion area as measured by radiographs was significantly lower in the fragmin- and K5-NSOS–treated groups than the vehicle group at sacrifice (P < 0.001; Fig. 3B and C). Histomorphometric examination revealed a significant reduction in tumor burden in bone in both heparin-like compound-treated groups as compared with the vehicle group (Fig. 3D and E). The total tumor area as determined per mouse was significantly smaller in both treatment groups than in the vehicle group (P < 0.05; Fig. 3E). Treatment with the heparin-like compounds did not have any significant effect on the size of the bone area or osteoclast number per mm tumor–bone interface as compared with the vehicle group (data not shown).

K5-NSOS and fragmin have different effects on human osteoclast activity in vitro

We also studied the effects of HLGAGs on human osteoclasts. Osteoclast precursors were cultured on bovine bone slices, and after the 7-day differentiation period, fragmin or K5-NSOS was added to the culture medium.
Fragmin did not affect osteoclast activity as determined by the resorption index CTX/TRACP 5b, whereas a dose-dependent inhibitory effect was observed with K5-NSOS (Fig. 4). K5-NSOS has lower anticoagulant activity than fragmin.

The applicability of heparin in cancer therapy is limited because of its anticoagulant activity. We compared the anticoagulant properties of K5-NSOS, fragmin, and heparin by measuring the antithrombin (anti-IIa) and antifactor Xa (anti-Xa) activity, which are specific coagulation parameters typically determined for heparin-like molecules. Fragmin and heparin showed the expected anti-IIa and anti-Xa activity. Only moderate anti-IIa and anti-Xa activity was observed with K5-NSOS. In addition, the effects of fragmin and K5-NSOS on APTT were tested as a function of concentration. K5-NSOS had clearly weaker prolonging effect on APTT than heparin. The concentration of K5-NSOS required to prolong APTT100s was almost equal to that of fragmin (Table 1).

Discussion

Our previous comparative genomic hybridization and genome-wide gene expression analyses indicated that the MDA-MB-231(SA) variant is closely related to the parental MDA-MB-231 cell line despite its remarkably enhanced bone metastatic propensity in vivo (6). Because TGF-β and IL-11 are important regulators of the metastatic process in the bone microenvironment and there is an increased TGF-β–induced IL-11 production in the MDA-MB-231(SA) as compared with the parental cells, we aimed to identify critical mediators of this induction among the genes that were highly overexpressed in the MDA-MB-231(SA) variant. One of the most highly overexpressed genes whose knockdown inhibited TGF-β–induced IL-11 production was HS6ST2. We subsequently showed that silencing of HS6ST2 inhibited Smad2/3/4-mediated transcriptional activity. The Smad pathway has previously been shown to be essential for IL-11 induction by TGF-β (13). HS6ST2 encodes an enzyme that attaches sulfate groups to glucosamine residues in heparan sulfate. Heparan sulfate glycosaminoglycans consist of a repeat disaccharide unit of either iduronic or glucuronic acid linked to a glucosamine. Distinct sulfation and acetylation patterns of O- and N-positions within the disaccharide units determine binding sites for numerous growth factors and cytokines (19, 20). Also TGF-β1 has been shown to bind to highly sulfated heparan sulfate glycosaminoglycans in vitro. Binding potentiates the biologic activity of TGF-β by protecting TGF-β1 from proteolytic degradation (17) and

![Figure 3.](image-url)
by preventing the formation of inactive complexes with α2-macroglobulin (15, 16). We found that silencing of enzymes that attach sulfate groups to C-6 of glucosamine residues, HS6ST1 and HS6ST2, inhibited TGF-β signaling. Indeed, our data showed that heparin and a chemically sulfated bacterial polysaccharide K5-NSOS with more than 90% sulfation degree at C-6 inhibited TGF-β-induced IL-11 production in MDA-MB-231(SA) cells. Fragmin did not show a similar effect which might be due to its lower molecular weight and consequently reduced interaction capacity.

Intracardiac injection of MDA-MB-231 human breast cancer cells into nude mice is a well-established model for breast cancer bone metastasis. We used this model and observed a significant inhibition of tumor growth in bone in the mice treated with K5-NSOS or fragmin. The anti-metastatic effects of these compounds were also observed as a reduction in tumor-induced cachexia and as an increase in body weight in the mice treated with HLGAGs. Tumor-induced bone destruction was significantly reduced by both HLGAGs. These data extend previous observations that have implicated antimetastatic effects for heparin and HLGAGs in animal model systems (21–26). Clinical studies have also indicated that heparin and HLGAGs may prolong survival of patients with cancer with solid tumors (3–5).

The complex coupling mechanisms between bone resorption and formation during the normal and tumor cell–induced bone remodeling make studies on the effects of HLGAGs on bone challenging. Furthermore, the mice used in this study were still growing and consequently had increased bone remodeling per se. Both fragmin and K5-NSOS reduced tumor-induced osteolysis as shown by the decreased lesion area in the X-ray radiographs. However, we did not see overall effect on total bone area. This could be due to the fact that the total bone area (both cortical and trabecular) was measured across the whole bone and not at the metaphysis resulting in dilution of the bone preservation effect of the drug that is noticeable particularly at the metaphysis. We also did not see a statistical

**Table 1. Coagulation activities of fragmin and K5-NSOS in vitro**

<table>
<thead>
<tr>
<th>Compound</th>
<th>Anti-Xa, IU/mg</th>
<th>Anti-IIa, IU/mg</th>
<th>APTT100s, s/g/mL</th>
</tr>
</thead>
<tbody>
<tr>
<td>K5-NSOS</td>
<td>14</td>
<td>6</td>
<td>10.9</td>
</tr>
<tr>
<td>Fragmin</td>
<td>155</td>
<td>74</td>
<td>9.4</td>
</tr>
<tr>
<td>Heparin</td>
<td>148</td>
<td>132</td>
<td>0.5</td>
</tr>
</tbody>
</table>

NOTE: The coagulation tests were conducted using the IL Test Heparin or the APTT Lyophilized Silica Kit. Anti-Xa and anti-IIa activities were determined by monitoring the level of paranitroaniline released from the chromogenic substrate in the enzyme reaction at 405 nm. APTT100s was determined by preparing dose–response curves as clotting time versus test compound concentration. The concentration required to prolong the coagulation time to 100s was estimated from the dose–response curve. All measurements were made according to the International Low-Molecular-Mass Heparin standard.
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